Vol.4., Issue.6., 2016 Nov-Dec.

REVIEW ARTICLE



ISSN: 2321-7758

LIGNOCELLULOSIC MATERIAL FOR PRODUCTION OF ETHANOL: A REVIEW

NIGUS WORKU^{1*}, HAILE HISHO²

^{1,2}Bule Hora University, Department of Chemical Engineering, Bule Hora, Ethiopia. *nigusu99@gmail.com



ABSTRACT

At present buildings with floating column is a typical feature in the modern Lignocellulosic biomass can be utilized to produce ethanol, a promising alternative energy source for the limited crude oil and climate change. The Present world energy scenario is focused at nonconventional sources. The biomass has emerged as one of the dependable nontraditional feed stocks for the production ethanol. Biomass derived from corn has become one of the primary feedstocks for bioethanol production for the past several years however, the argument of whether to use food as biofuel has led to a search for alternative non-food sources. Consequently, industrial research efforts have become more focused on low-cost large-scale processes for lignocellulosic feedstocks originating mainly from agricultural and forest residues along with herbaceous materials and municipal wastes. Although cellulosic-derived biofuel is a promising technology, there are some obstacles that interfere with bioconversion processes reaching optimal performance associated with minimal capital investment. This review summarizes current approaches on lignocellulosic-derived biofuel bioconversion and provides an overview on the major steps involved in cellulosic-based bioethanol processes. Key words: Lignocellulosic; Pretreatment; Hydrolysis; Fermentation; Ethanol

©KY PUBLICATIONS

1. INTRODUCTION

Global depletion of fossil fuels, rising fuel prices, environmental concerns, and pressures for oil independence are creating a strong market for bio-fuels [USDA, 2016]. Bio-fuels have the potential to be domestically and globally available for energy security, with most being carbon neutral (introducing no additional carbon to the global carbon cycle) or potentially carbon negative (if carbon coupled with sequestration) and supportable within the current agricultural infrastructure [DOE, 2015a]. Presently, one of the most promising alternatives for petro-fuels is bioethanol. Ethanol is a simple alkyl alcohol that can be used as a transport fuel in spark ignition engines. It has high octane levels and can be either blended into petrol or used in unmodified vehicles, or run as pure ethanol in a converted engine [Rudkin, E., 2002].

Ethanol can be produced from a variety of feed stocks, including sugar substances, such as sugarcane juice and molasses, as well as starchbased materials such as wheat and corn [Jones, A.M. *et.al, 1994*], where the corn starch based ethanol production is dominant in the US bioethanol industry[Korves, R, 2016]. However, this technology may not be practical in the long run due to the fact that wide planting of corn for ethanol production will compete for the finite arable land and thus threaten the national food security [Giampietro, M., *et al*, 1994]. Lignocellulosic biomass has been suggested as the most promising alternative for the traditional starch feedstock. Lignocellulosic feedstocks have the best well-towheel assessment, considering its abundance, low cost and high polysaccharides (cellulose and hemicellulose) content [Fujii, T *et al*, 2009].

Lignocellulosic biomass represents the major fraction of most plant matter. Common examples of lignocellulosic biomass include agricultural and forestry residues, the paper and much of the remaining organic fraction of municipal solid waste (MSW), industrial processing residues such as wastes in the paper and pulp industry, and herbaceous and woody plants grown as feedstocks for the production of fuels. The major fraction of such materials, typically of the order of 35-50%, is a polymer of glucose known as cellulose. The next largest fraction, of the order of 20-35%, is hemicellulose. Hemicellulose is also a polymer of sugars, but the types and distributions of these sugars vary depending upon the particular biomass source. For many types of lignocellulosic biomass, the five carbon sugar xylose represents the predominant fraction of the hemicellulose component. The third largest fraction, of the order of 15 to about 25%, is typically lignin, a phenylpropene polymer of complex composition that cannot be broken down to form sugar molecules. A number of other compounds such as plant oils, proteins, and ash make up the remaining fraction of the lignocellulosic biomass structure [Li, Xuan, 2010].

Biomass conversion involves three basic steps: (1) Pretreatment of raw feedstock to increase the accessibility of enzymes to the polysaccharides (cellulose and hemicellulose); (2) enzymatic hydrolysis to break down the lignocellulose constituents (polysaccharides) into a mixture of fermentable sugars; and (3) microbial fermentation, mediated by bacteria or yeast, to convert these sugars to ethanol [DOE, 2005b]

Ministry of Water and Energy (MoWE) of Ethiopia has developed bio-fuel strategy recently. The objective of the strategy is enhancing energy security and access to transport fuels. The program will also promote an agriculture based industry for increased agricultural and industrial outputs, employment and exports. The main goals of biofuel program in Ethiopia are (i) coordinating the production to gain 194.9 Million Liter at the end of strategic period, (ii) expanding bio-fuel blending facility through oil distribution companies, to have 8 benzene-ethanol blending stations at the end of the strategic period, (iii) enhancing benzeneethanol blending level to reach 25% and to use 64.4 million liter of bioethanol for blending, (iv) rising awareness and attract private investors on bio fuel development by hosting international forums and conferences, and (v) addition of 25 project designs on already existing 3 projects and coordinating biofuel project designs to reduce GHG emission and make the country beneficiary from CDM Program [Asrat G. W et al, 2015].

2. LIGNOCELLULOSIC FEEDSTOCK

2.1. Lignocellulosic Biomass Composition

Lignocellulosic material can generally be divided into three main components: cellulose (30-50%), hemicellulose (15-35%) and lignin (10-20%) [Pettersen RC, 1984]. Cellulose and hemicelluloses make up approximately 70% of the entire biomass and are tightly linked to the lignin component through covalent and hydrogenic bonds that make the structure highly robust and resistant to any treatment [Mielenz JR, 2001]. Potential lignocellulosic feedstocks and their composition are summarized in Table 1.

2.1.1. Cellulose

Cellulose is a structural linear component of a plant's cell wall consisting of a long-chain of glucose monomers linked b (1/4)- glycosidic bonds that can reach several thousand glucose units in length. The extensive hydrogen linkages among molecules lead to a crystalline and strong matrix structure [Ebringerova A et al, 2005]. This crosslinkage of numerous hydroxyl groups constitutes the microfibrils which give the molecule more strength and compactness. Although starchy materials require temperatures of only 60-70 $^{\circ}$ C to be converted from crystalline to amorphous texture, cellulose requires 320 $^{\circ}$ C as well as a pressure of 25 MPa to shift from a rigid crystalline

structure to an amorphous structure in water [Deguchi S, Et al, 206]. Cellulose is the most prevalent organic polymer and is approximately 30% of the plant composition [U.S., 2016]. Cotton, flax and chemical pulps represent the purest sources of cellulose (80-95% and 60-80%, respectively) while soft and hardwoods contain approximately 45% cellulose [Demirbas A, 2005].

2.1.2. Hemicellulose

Hemicellulose is an amorphous and variable structure formed of heteropolymers including hexoses (D-glucose, D-galactose and D-mannose) as well as pentose (D-xylose and L-arabinose) and may contain sugar acids (uronic acids) namely, D-glucuronic, D-galacturonic and methylgalacturonic acids [69]. Its backbone chain is primarily composed of xylan b (1/4)-linkages that include D-xylose (nearly 90%) and L-arabinose (approximately 10%) [Girio FM et al, 2010]. Branch frequencies vary depending on the nature and the source of feedstocks. The hemicelluloses of softwood are typically glucomannans while hardwood hemicellulose is more frequently composed of xylans [McMillan JD. 1993]

Although the most abundant component in hemicellulose, xylan composition still varies in each feedstock [Aspinall GO 19 80].

2.1.3. Lignin

Lignin is an aromatic and rigid biopolymer with a molecular weight of 10,000 Da bonded via covalent bonds to xylans (hemicelluloses portion) conferring rigidity and high level of compactness to the plant cell wall [Mielenz JR, 2001]. Lignin is composed of three phenolic monomers of phenyl propionic alcohol namely, coumaryl, coniferyl and sinapyl alcohol. Forest woody biomass is primarily composed of cellulose and lignin polymers. Softwood barks have the highest level of lignin (30-60%) followed by the hardwood barks (30-55%) while grasses and agricultural residues contain the lowest level of lignin (10-30% and 3-15%, respectively) [16]. Conversely, crop residues such as corn stover, rice and wheat straws are comprised mostly of a hemicellulosic heteropolymer that includes a large number of 5-carbon pentose sugars of primarily xylose [20]. Previously, little interest has been given to lignin chemistry potential on hydrolysis. However, lignin components are gaining importance because of their dilution effect on the process once solids are added to a fed batch hydrolytic or fermentation bioreactor in addition to their structure and concentration effects that would affect potential hydrolysis [21]. For instance, the adsorption of lignin to cellulases requires a higher enzyme loading because this binding generates a non-productive enzyme attachment and limits the accessibility of cellulose to cellulase [22]. Furthermore, phenolic groups are formed from the degradation of lignin. These components substantially deactivate cellulolytic enzymes and hence influence enzymatic hydrolysis. Chen et al. (2006) demonstrated that lignin modification via genetically engineering practices targeting its biosynthetic pathways could considerably reduce lignin formation and improve ethanol yield. However, this could be somewhat problematic as lignin components serve as the major plant defense system to pathogen and insects and its modification could disrupt the plants' natural protection[23].

PATHWAYS OF BIO-ETHANOL PRODUCTION

Lignocellulosic biomass be can transformed into bioethanol via two different approaches, (i.e. biochemical or thermochemicalbconversion) [24]. Both routes involve degradation of the recalcitrant cell wall structure of lignocellulose into fragments of lignin, hemicellulose and cellulose. Each polysaccharide is hydrolyzed into sugars that are converted into bioethanol subsequent followed by a purification process [25]. However, these conversion routes do not fundamentally follow similar techniques or pathways. The thermochemical process includes gasification of raw material at a high temperature of 800°C followed by a catalytic reaction. Application of high levels of heat converts raw material into synthesis gas (syngas) such as hydrogen, carbon monoxide and CO₂. In the presence of catalysts, the resulting syngas can be utilized by the microorganism Clostridium ljungdahlii to form ethanol and water can be further separated by distillation [26].

Unlike the thermochemical route, biochemical conversion involves physical (i.e. size reduction) or/and thermo-chemical with possible biological pretreatment. Biochemical pretreatment is mainly used to overcome recalcitrant material and increase surface area to optimize cellulose accessibility to cellulases [27]. The upstream operation is followed by enzymatic or acidic hydrolysis of cellulosic materials (cellulolysis) and conversion of hemicellulose into monomeric free sugars (saccharification) subsequent to biological fermentation where sugars are fermentedinto ethanol and then purified via distillation [25,27]. Concurrently, lignin, the most recalcitrant material of cell walls is combusted and converted into electricity and heat [28]. Overall, biochemical approaches include four unit-operations namely, pretreatment, hydrolysis, fermentation and distillation [29]. Currently the biochemical route is the most commonly used process [30].

3.1. Pretreatment Overview

Effective pretreatment is fundamental for optimal successful hydrolysis and downstream operations (See Fig.1) [31]. Pretreatment upstream operations include mainly physical, (i.e., biomass size reduction) and thermo chemical processes that involve the disruption of the recalcitrant material of the biomass. This upstream operation increases substrate porosity with lignin redistribution. Therefore, it enables maximal exposure of cellulases to cellulose surface area to reach an effective hydrolysis with minimal energy consumption and a maximal sugar recovery [27].

Zhu and Pan [32] concluded that the pretreatment process of woody biomass differs substantially from the agricultural biomass due to differences in their chemical composition and physical properties. Unlike woody biomass, agricultural residues pretreatment does not require as much energy as recalcitrant woody material to reach size reduction for further enzymatic saccharification. Emphasis on the importance of the energy consumption from the mechanical operation (size reduction) primarily based on the estimation of woody biomass pretreatment energy efficiency (Pretreatment ¼ Total sugar recovery (kg)/Total energy consumption (MJ)). In addition to sugar recovery and ethanol yield, this energy efficiency ratio and mass balance was deemed crucial for the complete estimation of pretreatment efficiency [32, 33]. Toxic inhibitory level estimation has also been considered important for evaluating pretreatment cost effectiveness primarily when dilute acid is added. Costly detoxification steps could be a major hindrance to reach highperformance pretreatment [34]. Overall, the ratio including energy consumption versus sugar yield with regard to feed stock versatility [32, 33] as well as toxic inhibitors formed per level of sugars recovered are of prime consideration on the estimation of the pretreatment efficiency and cost effectiveness of the operation in an effort to reach optimal conditions. Several pretreatment methods, namely, mechanical, chemical or microbiological have been used to remove the recalcitrant cell wall material of lignocellulosic biomass depending on the raw material being extracted [35]. More recently, there has been considerable advancement in development of pretreatment processes [36]. Table 2 illustrates some of the pretreatment methods that have been examined over the years. Although most of these treatments can liberate hemicellulose and cellulose from the cell wall, some of them remain economically unfeasible due to key technical issues. Furthermore, they are not all able to overcome the recalcitrant material found mainly in wood-based feedstocks. Typically, few treatments are endowed with ability to overcome feedstock versatility [37]. Unlike agriculture residues, forest and wood materials are high in lignin (approximately 29%) and cellulose (approximately 44%) [16] which renders them more recalcitrant. Agricultural residues such as corn stover, rice and wheat straws are mostly composed of hemicellulose (32%) and low levels of lignin (3-13%) conferring to them a less resistant texture but a higher level of pentose sugars rendering them less practical than woody recalcitrant material.

The most prevalent treatments include acid hydrolysis, hot water, dilute acid pretreatment and lime [92,93]. However, the conventional methods using acidic treatments (usually dilute sulfuric acid with concentrations below 4 wt% and temperatures greater than 160°C) [109] are always accompanied by formation of toxic inhibitors such as furfural from xylose and hydroxymethyl furfural (HMF) from glucose in addition to phenolics and acetic acid [93]. Acetic acid resulting from dilute acid pretreatment of agricultural residues as well as herbaceous and hardwoods is pH dependent and can reach a high concentration of approximately 10 g/L [36] that is more difficult to separate and detoxify than HMF and furfural. Unlike dilute acid pretreatment, ammonia fiber explosion (AFEX) treatments are sufficient to hydrolyze primarily agricultural residues such as corn stover and have not been associated with the formation of toxic products including HMF [97]. Given that woody feedstock is gaining increasing attention for its attractive attributes over low-lignin materials, organosolv along with steam explosion [38] and sulfite pretreatment to overcome recalcitrance (SPORL) [39] have become of prime interest for their ability to degrade high-lignin forest materials [32,39]. A recent study reported that steam explosion consumed the highest level of energy yielding the lowest pretreatment energy efficiency ratio of 0.26 kg sugar/MJ when compared to organosolv (0.31-0.40 kg sugar/MJ) and SPORL (0.35-043 kg sugar/MJ) [32]. While the organosolv treatments degrade high-lignin woody biomass including both softwood and hardwood, they produce considerable quantities of inhibitors namely furfural and HMF, yield a low hemicellulosic sugar concentration and are also associated with a high capital investment [40]. Consequently, SPORL remains the most attractive candidate for its flexibility and ability to overcome both hardwood and softwood recalcitrance with the highest sugar recovery and lowest energy consumption [32]

3.2. Hydrolysis

The success of the hydrolysis step is essential to the effectiveness of a pretreatment operation [28]. During this reaction, the released polymer sugars, cellulose and hemicellulose are hydrolyzed into free monomer molecules readily available for fermentation conversion to bioethanol [25]. There are two different types of hydrolysis processes that involve either acidic (sulfuric acid) or enzymatic reactions [41]. The acidic reaction can be divided into dilute or concentrated acid hydrolysis. Dilute hydrolysis (1-4%) requires a high temperature of 200-240 °C to disrupt cellulose crystals [42]. It is followed by hexose and pentose degradation and formation of high concentrations of toxic compounds including HMF and phenolics detrimental to an effective saccharification [43].

To optimize alcohol yield and reduce inhibitory and toxic byproducts the process uses sulfuric acid H_2SO_4 (0.5 wt%) that flows continuously to the biomass at a high temperature of 150-180 ^oC in a short period of time allowing for a greater sugar recovery [44]. Concentrated acid hydrolysis, the more prevalent method, has been considered to be the most practical approach [45]. Unlike dilute acid hydrolysis, concentrated acid hydrolysis is not followed by high concentrations of inhibitors and produces a high yield of free sugars (90%); however, it requires large quantities of acid as well as costly acid recycling, which makes it commercially less attractive [46]. While acid pretreatment results in a formation of reactive substrates when acid is used as a catalyst, acid hydrolysis causes significant chemical dehydration of the mono saccharides formed such that aldehydes and other types of degradation products are generated [43]. This particular issue has driven development of research to improve cellulolyticenzymes and enzymatic hydrolysis. Effective pretreatment is fundamental to a successful enzymatic hydrolysis [47].

During the pretreatment process, the lignocellulosic substrate enzymatic digestibility is improved with the increased porosity of the substrate and cellulose accessibility to cellulases. Trichoderma reesei is one of the most efficient and productive fungi used to produce industrial grade cellulolytic enzymes. The most common cellulase groups produced by T. reesei that cleave the b/1,4glycosidic bonds are b-glucosidase, endoglucanases and exoglucanases [40]. However, cellulase enzymes exposed to lignin and phenolic-derived lignin are subjected to adverse effects [48,49] and have demonstrated that phenolic derived lignin have the most inhibitory effects on cellulases. A study reported that a ratio of 4 mg to 1 mg peptides, reduced by half the concentration of cellulases (i.e. b-glucosidases) from T. reesei. This strain was also shown to be 10 to 10 fold more sensitive to phenolics than Aspergillus niger. In addition to phenolic components effect on Articles available online http://www.ijoer.in; editorijoer@gmail.com

cellulases, lignin has also an adverse effect on cellulases. As mentioned previously, the lignin adverse effect has two aspects including nonproductive adsorption and the limitation of the accessibility of cellulose to cellulase. Although considerable genetic modifications (GMs) have been deployed to transform lignin effects, lignin has been shown to be a potential source of self sustaining-energy and added-value components. Consequently, several research studies have determined practical approaches in eliminating inhibition of cellulases without involving GM approaches. Lui et al. [50] have demonstrated that the application of metal components namely, Ca(II) and Mg(II) via ligninemetal complexation substantially enhanced enzymatic hydrolysis. Additionally, Erickson et al. [51] have reported the importance of additives namely, surfactants and bovine serum albumin (BSA) in blocking lignin interaction with cellulases. Sewalt et al. [49] have reported that the adverse effect of lignin on cellulases can be surmounted by ammoniation and various N compounds. Moreover, the enzymatic treatment can be accomplished simultaneously with the engineered co-fermentation microbial process known as simultaneous saccharification and fermentation (SSF) [52]. This process has been of interest since the late 1970s for its effectiveness to minimize cellulolytic product inhibition and subsequently increase alcohol production [52]. Typically, separate hydrolysis and fermentation (SHF) processes involve the inhibition of the hydrolytic enzymes (cellulases) by saccharide products such as glucose and cellobiose. Unlike SHF, the SSF process combines hydrolysis and fermentation activities simultaneously and hence keeps the concentration of saccharides too low to cause any considerable cellulase inhibition [53].

3.3. Fermentation

Pretreatment and hydrolysis processes are designed to optimize the fermentation process . This natural, biological pathway depending on the conditions and raw material used requires the presence of microorganisms to ferment sugar into alcohol, lactic acid or other end products [25]. Moreover, industrial yeasts such as S. cerevisiae have been used in alcohol production mostly in the brewery and wine industries for thousands of years. S. cerevisiae has also been utilized for corn-based and sugar-based biofuel industries as the primary fermentative strain. Once becoming accessible for enzymatic or acidic hydrolysis, the pretreated cellulosic slurry is subsequently converted into fermentable free sugars. The sugars are mixed with water to form a broth. Typically, during batch fermentation S. cerevisiae ferments hexose sugars, mainly glucose, into ethanol in a large tank via the Embdene Meyerh of pathway under anaerobic conditions and controlled temperature. Yeastbased fermentation is always accompanied by formation of CO₂ by-products and supplemented by nitrogen to enhance the reaction. This conventional strain is optimal at a temperature of approximately 30[°]C and resists a high osmotic pressure in addition to its tolerance to low pH levels of 4.0 as well as inhibitory products [54]. S. cerevisiae can generate a high yield of ethanol (12.0-17.0% w/v; 90% of the theoretical) from hexose sugars [55].

Traditionally, separate hydrolysis and fermentation (SHF) sequential steps are used in bioethanol production. However, there is particular interest in targeting bioethanol production that can be derived from lignocellulosic biomass materials where both hexose and pentose sugars are available from the hemicellulose fraction. Despite its broad tolerance to stressful bioethanol process conditions, S. cerevisiae is not able to ferment sugars other than hexose. Unfortunately, lignocellulosic material includes a large proportion of hemicellulosic biomass that contains mainly pentose sugars such as D-xylose [56]. Moreover, an optimal fermentative microorganism should be tolerant to a high ethanol concentration and to chemical inhibitors formed during pretreatment and hydrolysis process. In response to this inability of S. cerevisiae to ferment pentose sugars, extensive efforts have been employed to develop genetically engineered microorganisms that are capable of fermenting pentose and hexose sugars simultaneously. An optimal fermentative microorganism should be able to utilize both hexose and pentose simultaneously with minimal toxic end-products formation. Different techniques including SSF and consolidated bioprocessing (CBP) Articles available online http://www.ijoer.in; editorijoer@gmail.com

have been developed to ensure the combination of hydrolysis (step 3) and fermentation (step 4) in one single reactor and thus, reduce product inhibition and operation costs. In addition to continuing downstream steps, CBP processing integrates both fermentation and cellulase formation in one fermentative/ cellulolytic microorganism [21]. However, despite the extensive range of prokaryotic and eukaryotic microorganisms that have been shown to be able to produce ethanol from sugars, most of them remain limited in terms of sugars co-fermentation, ethanol yield and tolerance to chemical inhibitors, high temperature and ethanol. In an effort to summarize relevant advantages and major limitations of microbial fermentative species.

3.4. Distillation

Bioethanol obtained from a fermentation conversion requires further separation and

purification of ethanol from water through a distillation process. Fractional distillation is a process implemented to separate ethanol from water based on their different volatilities. This process consists simply of boiling the ethanol water mixture. Because the boiling point of water $(100 \degree C)$ is higher than the ethanol-boiling point (78.3 ^oC), ethanol will be converted to steam before water. Thus, water can be separated via a condensation procedure and ethanol distillate recaptured at a concentration of 95% [23]. Typically, most largescale industries and biorefineries use a continuous distillation column system with multiple effects [57]. Liquid mixtures are heated and allowed to flow continuously all along the column. At the top of the column, volatiles are separated as a distillate and residue is recovered at the bottom of the column. Totally the general procedures in Fig. 2.

Table 1. The contents of cellulose, hemicellulose, and ligni	n in Lignocellulosic materials ^a	

Lignocellulosic materials	Cellullose (%)	Hemicellulose (%)	Lignin (%)
Hardwoods stems	40–55	24–40	18–25
Softwood stems	45–50	25–35	25–35
Nut shells	25–30	25–30	30–40
Corn cobs	45	35	15
Grasses	25–40	35–50	10–30
Wheat straw	30	50	15
Sorted refuse	60	20	20
Leaves	15–20	80–85	0
Cotton seed hairs	80–95	5–20	0
Newspaper	40–55	25–40	18–30
Waste papers from chemical pulps	60–70	10–20	5–10
Primary wastewater solids	8–15	NA ^b	24–29
Swine waste	6.0	28	NA ^b
Solid cattle manure	1.6–4.7	1.4–3.3	2.7–5.7
Coastal Bermuda grass	25	35.7	6.4
Switch grass	45	31.4	12.0

^a Source: Reshamwala et al. (1995), Cheung and Anderson (1997) and Boopathy (1998) ^bNA – not available. Articles available online http://www.ijoer.in; editorijoer@gmail.com

Table 2. Pre-treatment methods and key characteristics

Pretreatments	Key characteristics	
Dilute acid (H ₂ SO ₄ , HCL (0.5-	Practical and simple technique. Does not require thermal energy.	
5%)	Effective hydrolyze of hemicelluloses with high sugar yield, Generates toxic	
576)	inhibitors & Requires recovery steps	
Hot water	The majority of hemicelluloses can be dissolved.	
	4 No chemicals and toxic inhibitors.	
	Average solid load & Not successful with softwood.	
	High total sugar yield including pentose and hexose sugars.	
Lime	Effective against hardwood and agricultural residues.	
Lime	High pressure and temperature hinder chemical operation.	
	👃 Commercial scalability problem	
Ammonia fiber expansion	4 Effective against agricultural residues mainly corn stover without formation of	
	toxic end-products.	
(AFEX)	👃 Not suitable for high-lignin materials.	
	👃 Ammonia recovery	
	👃 No wastewaters	
Ammonia recycle	👃 High redistribution of lignin (85%)	
nerestation (ADD)	🖊 🛛 Recycling ammonia	
percolation (ARP)	👃 Theoretical yield is attained	
Steam explosion with	Effective against agricultural residues and hardwood.	
	High hemicelluloses fractions removal	
catalyst	A Not really effective with softwood	
Organosolv	+ High yield is enhanced by acid combination.	
-	Effective against both hardwood and softwood.	
	Low hemicellulosic sugar concentration	
	Formation of toxic inhibitors, Organic solvent requires recycling & High capital	
	investment	
Sulfite pretreatment top	Effective against high-lignin materials, both softwood and hardwood.	
	Highest pretreatment energy efficiency	
overcome	4 Minimum of inhibitors formation & Accommodate feedstocks versatility.	
recalcitrance (SPORL)	4 Steam explosion combined to SPORL in presence of catalyst becomes effective	
	against softwood materials	
	↓ Cost-effective.	
Ozone	Effectively remove lignin from a wide range of cellulosic material without	
	generating inhibitors.	
	🖕 Expensive	
Alkaline wet oxidation	4 The combination of oxygen, water, high temperature and alkali reduce toxic	
	inhibitors.	
	High delignification and solubilization of cellulosic material	
	Low hydrolysis of oligomers	
Fungal bioconversion	 Environmentally friendly 	
	Low use of energy and chemical	
	 Slow bioconversion 	

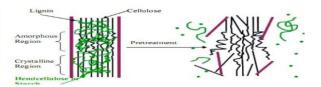
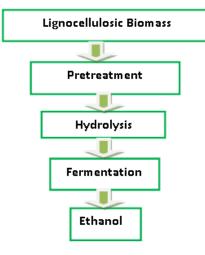


Figure 1. Schematic Representation on Biomass Pre-Treatment (Mosier et al, 2004)

International Journal of Engineering Research-Online

Vol.4., Issue.6., 2016 Nov-Dec.

Articles available online http://www.ijoer.in; editorijoer@gmail.com



A Peer Reviewed International Journal

Figure 2. Schematic diagram of cellulosic ethanol production

CONCLUSIONS

Ethanol is comes into traditional fuel for transportation in last decades and it is less polluting and clean burning fuel. Cellulosic-based ethanol is a potential alternative over food derived ethanol originating mainly from corn starch and sugarcane provided. This review highlights cellulosic biomass with different pretreatment, hydrolysis and fermentation methods to produce ethanol. The review assessed the potential of biomass for producing energy could increase into huge amount. Pretreatment, the most costly step is of particular concern due to the high recalcitrance of lignocellulosic raw materials. Given that lignocellulosic feedstock is a versatile material and bioethanol is a commodity product, it has been deemed imperative to design a general pretreatment combination that would be effective against a wide range of cellulosic material and hence deal with feedstock variability. For instance, researchers have shown that pretreatments involving steam explosion with either catalyst or lime are potential candidates to agricultural residues, herbaceous materials and hardwoods. The inability of steam explosion combined with catalyst to degrade softwood materials can be compensated by the low-cost and the energy efficient pretreatment approach. Genetic engineering also works to improve the efficiency of microorganisms for increase yield as well as minimum cost of production. Commonly Pichia stipites and saccharomyces cerevisiae is used by different researchers. Enzymatic hydrolysis may be most potent alternative process the for saccharification of complex polymer contains feed. Forest, industrial wastes typically unused and available in large amount. Agricultural waste is renewable, less costly and available in nature. Agriculture waste do not demand separate land, water and energy requirement. They do not have food value as well. Most of the municipal solid waste contains starch, lignocellulos, hemicelluloses and sugar that are sufficient for fermentation and very good raw materials compared to the other. While technological accomplishments and multiple research coalition efforts are still progressing, an efficient combination of the most advanced systems analysis and economical techniques designed to cope with feedstock versatility and commodity should emerge as the option of choice in an attempt to achieve optimal second-generation biofuel performance. In recent years it has been investigated that, instead of traditional feed stocks (starch crops), cellulosic biomass, including forest and industrial residues, agriculture waste and municipal waste, could be used as an ideally inexpensive and sufficient amount of sugar for production of ethanol by fermentation. Researchers mainly concern about cheapest and unused available resources to produce ethanol.

References

[1]. USDA ERS. Bioenergy. 2016

- [2]. DOE, Breaking the biological barriers to cellulosic ethanol: a joint research agenda. 2015a
- [3]. Rudkin, E. Bio-ethanol as a Transport Fuel. 2002
- [4]. Jones, A.M., Thomas, K.C., Ingledew, W.M.,. Ethanolic fermentation of blackstrap molasses and sugarcane juice using very high gravity technology. J. Agric. Food Chem. 1994 42 (5), 1242-1246.
- [5]. Korves, R., The potential role for corn ethanol in meeting the energy needs of the United States in. 2008 2016-2030.
- [6]. Giampietro, M., Ulgiati, S., Pimental, D.,.
 Feasibility of large-scale biofuel production.
 BioScience. 1997,47(9), 587-600
- [7]. Fujii, T., Fang, X., Inoue, H., Murakami, K., Sawayama, S.,. Enzymatic hydrolyzing performance of *Acremonium cellulolyticus* and *Trichoderma reesei* against threelignocellulosic materials. Biotechnol. Biofuels 2009, 2, 24.
- [8]. Li, Xuan, "Bioethanol production from lignocellulosic feedstock using aqueous ammonia pretreatment and simultaneous saccharification and fermentation (SSF): process development and optimization" (2010).
- [9]. DOE, Making bioethanol cost competitive 2005b.
- [10]. Asrat G. W, Belay W. and Bhagwan S. C. Theoretical Estimation of Bio-Ethanol Potential of Wet Coffee Processing Waste (Pulp Juice and Mucilage). International Journal of Engineering Innovation & Research 2015;42: 2243 – 2247
- [11]. Pettersen RC. The chemical composition of wood. In: Rowell RM, editor. The chemistry of solid wood. Advances in chemistry series, vol. 207. Washington, DC: American Chemical Society; 1984. p. 115-6.
- [12]. Mielenz JR. Ethanol production from biomass: technology and commercialization status. Curr Opin Microbiol 2001;4:324-5.
- [13]. Ebringerova A, Hromadkova Z, Heinze T. Hemicellulose Adv Polym Sci 2005; 186:1-67.

- [14]. Deguchi S, Mukai SA, Tsudome M, Horikoshi K. Facile generation of fullerene nanoparticles by hand-grinding. Adv Mater 2006;18:729-32.
- [15]. U.S. Department of Energy Biomass Program. 2016.
- [16]. Demirbas A. Bioethanol from cellulosic materials: a renewable motor fuel from biomass. Energy Source 2005;27:327-37.
- [17]. Girio FM, Fonseca C, Carvalheiro F, Duarte LC, Marques S, Bogel-Lukasic R. Hemicellulose Bioresour Technol 2010;101:4775-800.
- [18]. McMillan JD. Pretreatment of lignocellulosic biomass. In: Himmel ME, Baker JO, Overend RP, editors. Enzymatic conversion of biomass for fuel production. Washington, D.C: American Chemical Society; 1993. p. 292-323.
- [19]. Aspinall GO. Chemistry of cell wall polysaccharides. In: Preiss J, editor. The biochemistry of plants (a comprehensive treatise). Carbohydrates structure and function, vol. 3. NY: Academic Press; 1980. p. 473-500.
- [20]. Foody BE, Foody KJ. Development of an integrated system for producing ethanol from biomass. In: Klass DL, editor. Energy from biomass and waste. Chicago: Institute of Gas Technology; 1991. p. 1225-43.
- [21]. Ladisch MR, Mosier NS, Kim Y, Ximenes E, Hogsett D. Converting cellulose to biofuels. SBE special supplement biofuels. CEP 2010;106(3):56-63.
- [22]. Chen F, Srinivasa RMS, Temple S, Jackson L, Shadle G, Dixon RA. Multi-site genetic modulation of monolignol biosynthesis suggests new routes for formation of of syringyl lignin and wall-bound ferulic acid in alfalfa (Medicago sativa L.). Plant J 2006;48:113-24.
- [23]. Li X, Weng JK, Chapple C. Improvement of biomass through lignin modification. Plant J 2008;54:569-81.
- [24]. Demirbas A. Progress and recent trends in biofuels. Prog Energy Combust Sci 2007;33:1-18.

- [25]. Chandel AK, Chan E, Rudravaram R, Narasu ML, Rao LV, Ravindra P. Economics and environmental impact of bioethanol production technologies: an appraisal. Biotechnol Mol Biol Rev 2007;2:14-32.
- [26]. Mu D, Seager T, Suresh Rao P, Zhao F. Comparative life cycle assessment of lignocellulosic ethanol production: biochemical versus thermochemical conversion. Environ Manage 2010;46:565-78.
- [27]. Yang B, Wyman CE. The key to unlocking low-cost cellulosic ethanol. Biofuels Bioprod Bioref 2008;2:26-40.
- [28]. Gamage J, Howard L, Zisheng Z. Bioethanol production from lignocellulosic biomass. J Biobased Mater Bioenerg 2010;4:3-11.
- [29]. Sánchez ÓJ, Cardona CA. Trends in biotechnological production of fuel ethanol from different feedstocks. Bioresour Technol 2008; 99:5270-95.
- [30]. Fehrenbacher K. Logen suspends U.S. cellulosic ethanol plant plans [accessed 2016.
- [31]. Wyman CE. What is (and is not) vital to hydrolysate to ethanol. In: Himmel ME, Baker JO, Overend RP, editors. Enzymatic conversion of biomass for fuels production. ACS symposium series, vol. 566. Washington, DC: American Chemical Society; 1994. p. 411-37.
- [32]. Zhu JY, Pan HJ. Woody biomass treatment for cellulosic ethanol production: technology and energy consumption evaluation. Bioresour Technol 2010; 101:4992-5002.
- [33]. Zhu JY, Zhu W, OBryan P, Dien BS, Tian S, Gleisner R, et al. Ethanol production from SPORL-pretreated lodge pole pine: Preliminary evaluation of mass balance and process energy efficiency. Appl Microbiol Biotechnol 2010;86: 1355-65.
- [34]. Laser M, Schulman D, Allen SG, Lichwa J, Antal MJ, Lynd LR. A comparison of liquid hot water and steam pretreatments of sugar cane bagasse for bioconversion to ethanol. Bioresour Technol 2002;81:33-44.

- [35]. Wyman CE, Dale BE, Elander RT, Holtzapple M, Ladisch MR, Lee YY. Coordinated development of leading biomass pretreatment technologies. Bioresour Technol 2005;96:1959-66.
- [36]. Banerjee S, Mudliar S, Sen R, Balendu BG, Chakrabarti T, Pandy RA. Commercializing lignocellulosic bioethanol: technology bottlenecks and possible remedies. Biofuel Bioprod Biorg 2009;4:77-93.
- [37]. Klinke HB, Thomsen AB, Ahring BK. Inhibition of ethanol-producing yeast and bacteria by degradation products produced during pretreatment of biomass. Appl Microbiol Biotechnol 2004;66:10-26.
- [38]. Ohgren K, Bura R, Saddler J, Zacchi G. Effect of hemicellulose and lignin removal on enzymatic hydrolysis of steam pretreated corn stover. Bioresour Technol 2007 ;98(13):2503-10.
- [39]. Zhu JY, Pan XJ, Wang GS, Gleisner R. Sulfite pretreatment (SPORL) for robust enzymatic saccharification of spruce and red pine. Bioresour Technol 2009; 100:2411-8.
- [40]. Eggeman T, Elander RT. Process and economic analysis of pretreatments technologies. Bioresour Technol 2005;96:2019-25.
- [41]. Nazhad MM, Ramos LP, Paszner L, Saddler JN. Structural constraints affecting the initial enzymatic hydrolysis of recycled paper. Enzyme Microbiol Technol 1995;17:66-74.
- [42]. Xiang Q, Lee YY, Pettersson PO, Torget RW. Heterogeneous aspects of acid hydrolysis of a-cellulose. Appl Biochem Biotechnol 2003;105/108:505-14.
- [43]. Sun Y, Cheng J. Hydrolysis of lignocellulosic materials for ethanol production: a review. Bioresour Technol 2002;83:1-11.
- [44]. Harris EE, Beglinger E. The Madison woodsugar process. Report no R1617. Madison, Wisconsin: US Department of Agriculture, Forest Products Laboratory; 1946
- [45]. Torget R, Walter P, Himmel M, Grohmann K. Dilute acid pretreatment of short rotation woody and herbaceous crops. Appl Biochem Biotechnol 1991; 24-25:115-26.

- [46]. Hamelinck CN, Hooijdonk G, Faaij APC. Ethanol from lignocellulosic biomass: techno-economic performance in short-, middle- and long-term. Biomass Bioenerg 2005;28:384-410.
- [47]. Cardona CA, Quintero JA, Paz IC. Production of bioethanol from sugarcane bagasse: status and perspectives. Bioresour Technol 2009;101:4754-66.
- [48]. Larsson S, Palmqvist E, Hahn-Hagerdal B, Tengborg C, Stenberg K, Zacchi G, et al. The generation of fermentation inhibitors during dilute acid hydrolysis of softwood. Enzyme Microbiol Technol 1999;24:151-9.
- [49]. Boone RS, Kozlik CJ, Bois CJPJ, Wengert PPJEM. Dry kiln schedules for commercial woods temperate and tropical. Gen. Tech. Rep. FPL_GTR_57. Madison, WI: U.S. Department of Agriculture, Forest Service, Forest Products Laboratory; 1988.
- [50]. Markwardt LJ, Wilson TRC. Strength and related properties of woods grown in the United States. Tech. Bull. 479. Washington, DC: U.S. Department of Agriculture, Forest Service. U.S. Government Printing Office; 1935
- [51]. Erickson T, Borjesson J, Tjerneld F. Mechanism of surfactant effect in enzymatic hydrolysis of lignocelluloses. Enzyme Microbiol Technol 2002;31: 353-64.
- [52]. Bisaria VS, Ghose TK. Biodegradation of cellulosic materials: substrate, microorganisms, enzymes and products. Enzyme Microbiol Technol 1981;3: 90-104.
- [53]. Zhu JY, Pan HJ. Woody biomass treatment for cellulosic ethanol production: technology and energy consumption evaluation. Bioresour Technol 2010; 101:4992-5002
- [54]. Hahn-Hägerdal B, Karhumaa HBK, Fonseca C, Spencer-Martins I, Gorwa-Grauslund MF. Toward industrial pentose-fermenting yeast strains. Appl Microbiol Biotechnol 2007;74:937-53.
- [55]. Claassen PAM, Lopez C, Sijtsma AM, Weusthuis L, Van Lier RA, Van Niel JB, et al. Utilization of biomass for the supply of

energy carriers. Appl Microbiol Biotechnol 1999;52:741-55.

- [56]. Martin C, Galbe Wahlbom M, Hagerdal BH, Jonsson JL.Ethanol production from enzymatic hydrolysates of sugarcane bagasses using recombinant xylose e utilizing Saccharomyces cerevisisae. Enzyme Microbiol Technol 2002;31(2):274-82.
- [57]. Kent NL, Evers AD. Malting, brewing and distilling. In: Kent's technology of cereals.4th ed., vol. 4. Cambridge: Woodhead Publishing; 1994. p. 218-32.